

FORM PTO-1390

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2577-124A

**TRANSMITTAL LETTER TO THE UNITED STATES  
DESIGNATED/ELECTED OFFICE (DO/EO/US)  
CONCERNING A FILING UNDER 35 U.S.C. 371**

U.S. Application No. (if known, see 37 CFR 1.5)

10/009590

INTERNATIONAL APPLICATION NO.  
PCT/SG99/00058

INTERNATIONAL FILING DATE  
June 11, 1999

PRIORITY DATE CLAIMED  
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TITLE OF INVENTION HIGH-EFFICIENCY AGROBACTERIUM-MEDIATED TRANSFORMATION OF COTTON  
USING PETIOLE EXPLANTS

APPLICANT(S) FOR DO/EO/US Zhi Xian CHEN and Lian Hui ZHANG

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☐ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below.
4. ☒ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
  - a. ☒ is attached hereto (required only if not communicated by the International Bureau).
  - b. ☐ has been communicated by the International Bureau.
  - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US)
6. ☐ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
  - a. ☐ is attached hereto.
  - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
  - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
  - b. ☐ have been communicated by the International Bureau.
  - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
  - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

**ITEMS 11. TO 20. below concern other document(s) or information included:**

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☐ A **FIRST** preliminary amendment.
14. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
15. ☐ A substitute specification.
16. ☐ A change of power of attorney and/or address letter.
17. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821-1.825
18. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4).
19. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
20. ☒ Other items or information: One drawing sheet (Fig. 1); a copy of the International Preliminary Examination Report; a copy of the International Search Report.

U.S. APPLICATION NO. 10/009590

INTERNATIONAL APPLICATION NO.  
PCT/SG99/00058

ATTORNEY DOCKET NO.  
2577-124

17. [X] The following fees are submitted:

**Basic National Fee (37 CFR 1.492)(a)(1)-(5):**

Neither international preliminary examination fee (37 CFR 1.482)  
nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and  
International Search Report Not Prepared by EPO or JPO. .... \$ 1,040.00  
International preliminary examination fee (37 CFR 1.482) not paid to USPTO  
but International Search Report has been prepared by the EPO or JPO ..... \$ 890.00  
International preliminary examination fee (37 CFR 1.482) not paid to USPTO  
but international search fee (37 CFR 1.445(a)(2)) paid to USPTO ..... \$ 740.00  
International preliminary examination fee (37 CFR 1.482) paid to USPTO  
but claims did not satisfy provisions of PCT Article 33(1)-(4) ..... \$ 710.00  
International preliminary examination fee (37 CFR 1.482) paid to USPTO  
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Surcharge of \$130.00 for furnishing the oath or declaration later than [ ] 20 [ ] 30  
months from the earliest claimed priority date (37 CFR 1.492(e)).

\$

Claims

Number Filed

Number Extra

Rate

Total Claims

18 -20 =

X \$18.00

\$

Independent Claims

1 - 3 =

X \$84.00

\$

Multiple dependent claim(s) (if applicable) NONE

+ \$280.00

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**TOTAL OF ABOVE CALCULATIONS =**

\$ 890.00

☐ Applicant claims small entity status. See 37 CFR 1.27. The fees indicated  
above are reduced by 1/2.

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**SUBTOTAL =**

\$ 890.00

Processing fee of \$130.00 for furnishing the English translation later than [ ] 20 [ ] 30  
months from the earliest claimed priority date (37 CFR 1.492(f)).

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**TOTAL NATIONAL FEE =**

\$ 890.00

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be  
accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property

\$

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Amount to be  
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- a. ☒ A check in the amount of \$ 890.00 to cover the above fees is enclosed.
- b. ☐ Please charge my Deposit Account No. 02-2135 in the amount of \$ to cover the above fees.  
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**NOTE:** Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a)  
or (b)) must be filed and granted to restore the application to pending status.

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Registration Number

1 / PARTS

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Technical Field

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## Background

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Tissue Culture of Cotton: In 1935, Skovsted reported the first embryo culture of cotton. Beasley

(1971) reported callus formation in cotton as an outgrowth from the micropylar end of fertilized ovules on Murashige & Skoog (MS) medium. Somatic embryogenesis was achieved from a suspension culture of *G. klotzschianum* (Price & Smith, 1979). In 1983, Davidonis & Hamilton first succeeded in efficient and repeatable regeneration of cotton (*G. hirsutum* L.) plants from callus after two-year cultivation. Cotton plants were since regenerated through somatic embryogenesis from different explants (Zhang & Feng, 1992; Zhang, 1994) including cotyledon (Davidonis et al., 1987; Davidonis & Hamilton, 1983; Finer, 1988; Firoozabady et al., 1987), hypocotyl (Cousins et al., 1991; Rangan & Zavala, 1984; Rangan & Rajasekaran, 1996; Trolinder & Goodin, 1988; Umbeck et al., 1987, 1989), stem (Altman et al., 1990; Bajaj et al., 1989; Chen, et al. 1987; Finer & Smith, 1984), shoot apex (Bajaj et al., 1985; Gould et al., 1991; Turaev & Shamina, 1986), immature embryo (Beasley, 1971; Stewart & Hsu, 1977, 1978), petiole (Finer & Smith, 1984; Gawel et al., 1986; Gawel & Robacker, 1990), leaf (Finer & Smith, 1984; Gawel & Robacker, 1986), root (Chen & Xia, 1991; Kuo et al., 1989), callus (Finer & McMullen, 1990; Trolinder et al., 1991) and protoplast (Chen et al., 1989).

Transformation of cotton: *Agrobacterium*-mediated cotton transformation was first reported a decade ago with hypocotyl and cotyledon as explants (Firoozababy et al., 1987; Umbeck et al., 1987). Several useful genes have been introduced into cotton via *Agrobacterium*-mediated transformation, including insect and herbicide resistance genes (Perlak et al., 1990; Trolinder et al., 1991; Chen et al., 1994). Explants

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(such as hypocotyl, cotyledon, callus generated from hypocotyl and cotyledon, as well as immature embryos) have been used for *Agrobacterium*-mediated transformation and particle bombardment (de Framond et al., 1983; Finer & McMullen, 1990; Firoozabady et al., 1987; Perlak et al., 1990; Rangan & Rajasekaran, 1996; Rajasekaran et al., 1996; Trolinder et al., 1991; Umbeck et al., 1987, 1989, 1992). In addition, meristematic tissue of excised embryonic axes has also been used for cotton transformation by particle bombardment (Chlan et al., 1995; John, 1996; John & Keller, 1996; McCabe & Martinell, 1993). Zhou et al. (1983) transformed cotton by injecting DNA into the axile placenta one day after self-pollination.

However, the transformation rates were generally low, ranging from 20 to 30% when hypocotyl were used as explant (Firoozababy et al., 1987; Cousins et al., 1991; Rajasekaran et al., 1996). A significantly higher transformation efficiency, up to 80%, was reported when cotyledon was used as explant and the *ocs* gene encoding octopine synthetase used as the reporter gene (Firoozababy et al., 1987). However, the validity of octopine as a marker for transformation is questionable because octopine has been found in several plant species certainly not transformed by infection with *A. tumefaciens* (Wendt-Gallitelli and Dobrigkeit, 1973). A more recent report indicated that the transformation efficiency of cotyledon was about 20 to 30% (Cousins et al., 1991). The transformation efficiency was even lower when particle bombardment method was used (Keller et al., 1997). A difference in the type of explants used for transformation could have a significant effect on the efficiency of

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transformation and regeneration. It has been reported, for example, that for reducing false positive transformants, cotyledon was a better explant than hypocotyledon (Firoozabady et al., 1987).

5 Cotton transformation also is highly dependent on genotype (Trolinder, 1985a, 1986; Trolinder & Goodin, 1987, 1988a, 1988b; Trolinder & Chen, 1989). Apart from a few cultivars which are regenerable and transformable, such as *Gossypium hirsutum* cv. Coker 312  
10 and *G. hirsutum* Jin 7, most other important elite commercial cultivars, such as *G. hirsutum* cv. D&P 5415 and *G. hirsutum* cv. Zhongmian 12, are not regeneratable and transformable by these methods. The absence of a high-efficiency plant regeneration method has been  
15 regarded as a major obstacle to the application of *Agrobacterium*-mediated transformation to cotton (Gawel et al., 1986; Firoozabady et al., 1987).

#### Summary of the Invention

To overcome the problems associated with  
20 previously reported methods, an efficient transformation procedure using petiole as an explant has been developed, along with a set of correspondingly improved media. This method provides several advantages in comparison to the hypocotyl and cotyledon  
25 methods: (1) explants are easy to obtain; (2) transformation efficiency is higher; (3) *Agrobacterium* contamination is very rare; (4) efficiency in regeneration is higher; and (5) the time from transformation to regeneration of plantlets is reduced.  
30 Two cotton varieties, i.e. Coker 312 and Si-Mian 3, have been successfully transformed with this method, and more than 30 independent transgenic lines from

Coker 312 showing strong activity of the marker transgene have been obtained. This method is applicable to other cotton varieties such as Jin 7 and Ji 713 from China, Siokra 1-3 from Australia, T25, Coker 201 and Coker 310 from the U.S.A.

#### Brief Description of the Figure

Figure 1 shows the plasmid pBI121GFP, containing GFP as the reporter gene and the NPT II (neomycin phosphotransferase) gene as a selectable marker, used for *Agrobacterium*-mediated transformation of cotton petiole according to the methods of the present invention.

#### Detailed Description

An efficient method is disclosed for genetic transformation of cotton plants, including elite lines, using cotton petiole as an explant. By using petiole explants, plus a set of improved media, transformation efficiency is significantly enhanced and the time required from transformation to regeneration is shortened in comparison to previously reported methods.

By using the methods of the present invention, the whole process from *Agrobacterium* transformation to the regeneration of transgenic plantlets can take about 6-7 months. The reported hypocotyl and cotyledon methods usually required 7-9 months or longer to complete the same process (Cousins et al., 1991; Chen et al., unreported observation). Another two months were required for growing the small plantlets to a suitable size for potting in soil.

Techniques for introducing exogenous genes into *Agrobacterium* such that they will be transferred stably

to a plant or plant tissue exposed to the *Agrobacterium* are well-known in the art and do not form part of the present invention. It is advantageous to use a so-called "disarmed" strain of *Agrobacterium* or Ti

5 plasmid, that is, a strain or plasmid wherein the genes responsible for the formation of the tumor characteristic of the crown gall disease caused by wild-type *Agrobacterium* are removed or deactivated. Numerous examples of disarmed *Agrobacterium* strains are  
10 found in the literature (e.g., pAL4404, pEHA101 and pEH105 (Walkerpeach & Veltern, 1994)). It is further advantageous to use a so-called binary vector system, such as that described in Schilperoort et al., 1990, 1995. A binary vector system allows for manipulation  
15 in *E. coli* of the plasmid carrying the exogenous gene to be introduced into the plant, making the process of vector construction much easier to carry out.

Similarly, vector construction, including the construction of chimeric genes comprising the exogenous  
20 gene that one desires to introduce into the plant, can be carried out using techniques well-known in the art and does not form part of the present invention.

Chimeric genes should comprise promoters that have activity in the host in which expression is desired.

25 For example, it is advantageous to have a series of selectable markers for selection of transformed cells at various stages in the transformation process. A selectable marker (for example a gene conferring resistance to an antibiotic such as kanamycin,  
30 cefotaxime or streptomycin) linked to a promoter active in bacteria would permit selection of bacteria containing the marker (i.e., transformants). Another selectable marker linked to a plant-active promoter,



such as the CaMV 35S promoter or a T-DNA promoter such as the NPT II NOS promoter, would allow selection of transformed plant cells. The exogenous gene that is desired to be introduced into the plant cell should  
5 comprise a plant-active promoter in functional relation to the coding sequence, so that the promoter drives expression of the gene in the transformed plant. Again, plant-active promoters, such as the CaMV 35S, the NPT II NOS promoter or any of a number of tissue-specific promoters, are well-known in the art and  
10 selection of an appropriate promoter is well within the ordinary skill in the art.

The present method can be used to produce transgenic plants expressing any number of exogenous  
15 genes, and is not limited by the choice of such a gene. The selection of the desired exogenous gene depends on the goal of the researcher, and numerous examples of desirable genes that could be used with the present invention are known in the art (e.g., the family of  
20 *Bacillus thuringiensis* toxin genes, herbicide resistance genes such as shikimate synthase genes that confer glyphosate resistance, U.S. Patent No. 5,188,642, or a 2,4-D monooxygenase gene that confers resistance to 2,4-dichlorophenoxyacetic acid (2,4-D),  
25 Bayley et al., Theoretical and Applied Genetics, vol. 82, pp. 645-49, male sterility genes such as the antisense genes of U.S. Patent No. 5,741,684 (Fabijanski, et al.), or even the elaborate crop protection systems described in U.S. Patent No.  
30 5,723,765 (Oliver et al.)).

Cotton regeneration is considered in the art to be heavily variety-dependant. The Coker series of cotton varieties have been shown to be relatively easy to

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transform. However, DP 5412, Zhongmain 12 and many other varieties still have difficulties associated with regeneration. The situation is the same for *G. barbadense* and other diploid species. While somatic embryogenesis and regeneration of whole plants is a highly genotype-dependent process in cotton, successful transformation and regeneration of two distinct cotton varieties, i.e. Coker 312 from U.S.A. and Si-Mian 3 from China, has been demonstrated using the methods of the present invention. It is therefore believed that the present invention has wide applicability to transformation of a variety of cotton lines.

Transgene integration in the genome of cotton produced by the methods of the present invention was confirmed using standard Southern hybridization techniques, as can identification of the copy number of the inserted transgene in each transgenic line (see Example 6, below). The F1 generation of transgenic cotton can be tested for the presence of the transgene, and inheritance pattern of the transgene in the F1 generation can be analyzed to confirm stability and inheritability.

As compared with other reported protocols, the cotton transformation system of the present invention has higher transformation efficiency and survival rate. This is attributable to several factors. In the present invention, petiole was used as an explant for transformation. Different types of cotton explants can have significant effects on the efficiencies of plant transformation and regeneration (Firoozabady et al., 1987). Induction of somatic embryogenesis from petiole was reported previously. But regeneration was either unsuccessful or very poor (Finer and Smith, 1984; Gawel

et al., 1986). With the present invention, the efficiency of regeneration was significantly improved by using the improved media discussed below. In a preferred embodiment, calli of high quality were  
5 obtained when tender petioles rich in parenchyma cell in primary vascular bundle tissue were cultured in the MMSI medium (described below) with low concentrations of 2,4-D and Kinetin.

With the present invention, the time for embryo  
10 induction in suspension culture can be shortened to 10 - 14 days, from a previously reported 3 weeks (Cousins et al., 1991). It was found that a shortened period of suspension culture treatment is important for high frequency induction of embryogenesis. It is also  
15 important for reducing production of abnormal embryos, since a high percentage of vitreous embryos that are poor in regeneration are produced when cotton calli are maintained in suspension culture for too long (Chen et al, unpublished observation).

For maximum cell growth at different stages except  
20 at the young plant growing stage, glucose was used as the sole carbon source. The amount of glucose in the media can be from about 10 to about 50 g/l, preferably about 30 g/l. At the young plant growing stage,  
25 glucose and sucrose at about 10 g/l respectively as carbon sources are preferable for promotion of healthy plantlets growth.

For growth of callus, embryogenesis and callus proliferation, pH range can be from 5.8 to 7.5,  
30 preferably pH 6.2 - 7.0, most preferably at pH 6.5. A medium of pH 7.0 is preferable for healthy root growth of plantlets.

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For effective callus initiation and induction of the potency of embryogenesis, low concentrations of 2,4-D and kinetin in the callus induction and selection medium is important. The amount of 2,4-D can be from 0 to about 0.5 mg/l, preferably about 0.05 mg/l. The amount of kinetin can be from 0.0 mg/l to about 1.0 mg/l, preferably about 0.1 mg/l. In the callus differentiation stage and embryoid germination stage, best result were obtained when no plant hormone was added to the media.

The amino acids asparagine and glutamine are better nitrogen sources than inorganic ammonia nitrogen for specifically supporting embryoids germination and root development. In the embryoid germination medium, the amount of asparagine can be about 200 to about 1000 mg/l, preferably about 500 mg/l. The amount of glutamine can be about 500 to about 2000 mg/l, preferably about 1000 mg/l. With these optimized nitrogen sources, the growth of non-embryogenic calli was inhibited while the germination, growth and root development of embryoids were preferentially promoted.

At different stages of cotton transformation except co-culture with *Agrobacterium*, plant tissue and callus are preferably maintained at 28°C but can be varied from 25-35°C. For effective transformation, temperature in co-culture stage should not be higher than 28°C. A light condition of 16 hrs. light (60-90,  $\mu\text{Em}^{-2}\text{S}^{-1}$ ) and 8 hrs. dark per day is preferable for all stages of cotton transformation and regeneration.

Unlike previously reported transformation and regeneration protocols (Umbeck et al., 1987; Firoozabady et al., 1987, Cousins et al.), the media used in the present invention are optimized in several

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respects: (a) glucose is used as a sole carbon source in all culture media except in the medium used to culture young plants previous to planting out in the greenhouse; (b) the media is adjusted to higher pH value (6.5-7.0); (c) lower concentration of 2,4-D (0.05mg/l) and kinetin (0.1 mg/l) is used only at callus initiation stage, no hormone is used at other stages; (d) asparagine and glutamine are used to replace inorganic ammoniac nitrogen in the medium used for embryoid germination. These modifications are adapted for the physiological requirement of cotton embryoid development and plantlet growth. It has been found that healthy embryoid development and plantlet growth, especially root system development, are largely attributable to these optimized media. For example, it has been found that asparagine and glutamine were better nitrogen source than inorganic ammonia nitrogen for supporting embryoid germination and root development. In the preferred MMS3 medium (described below), which contains asparagine and glutamine as the nitrogen source, the growth of non-embryogenic calli was inhibited while the germination, growth and root development of embryoids were preferentially promoted. Because of the healthy root development, the survival rate of potted transgenic cotton plants obtained by the methods of the present invention is almost 100%. With the reported hypocotyl and cotyledon protocols (Umbeck et al., 1987; Firoozabady et al., 1987), poor root development has been regarded as the main reason accounting for poor survival rate of potted transgenic cotton plants.

The following are preferred plant tissue culture media used in the Examples:

- (1) Seedling growing medium (per liter):
- 5         $\frac{1}{2}$  MS basal salt mixture (Sigma M5524)  
          0.9 g  $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$   
          2.0 g gellan gum (Phytigel™, Sigma)  
          pH 7.0
- (2) Petiole pre-culture medium (per liter):
- 10       MS basal salt mixture  
          0.9 g  $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$   
          2.0 g gellan gum (Phytigel™, Sigma)  
          pH 7.0
- (3) Co-culture medium (per liter):
- 15       MS basal salt mixture  
          10 mg Thiamine-HCl  
          1 mg Pyridoxine-HCl  
          1 mg Nicotinic acid  
          100 mg Myo-inositol  
          0.05 mg 2,4-dichlorophenoxyacetic acid (2,4-D)
- 20       0.1 mg Kinetin  
          30 g Glucose  
          0.9 g  $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$   
          2.0 g gellan gum (Phytigel™, Sigma)  
          pH 6.5
- 25       (4) MMS1 - callus induction and selection medium (per  
          liter):  
          Co-culture medium  
          50 mg Kanamycin  
          500 mg Cefotaxime

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## (5) MMS2 - differentiation medium (per litre):

MS basal salt mixture  
 10 mg Thiamine-HCl  
 1 mg Pyridoxine-HCl  
 5 1 mg Nicotinic acid  
 100 mg Myo-inositol  
 1.9 g KNO<sub>3</sub>  
 30 g Glucose  
 0.9 g MgCl<sub>2</sub>·6H<sub>2</sub>O  
 10 2.0 g gellan gum (Phytigel™, Sigma)  
 pH 6.5

## (5) MMS3 - embryoid germination medium (per litre):

3.8 g KNO<sub>3</sub>  
 440 mg CaCl<sub>2</sub>·H<sub>2</sub>O  
 15 375 mg MgSO<sub>4</sub>·7H<sub>2</sub>O  
 170 mg KH<sub>2</sub>PO<sub>4</sub>  
 1 g Glutamine  
 500 mg Asparagine  
 43 mg EDTA ferric-Na salt  
 20 MS micronutrients (Murashige and Skoog, 1962)  
 10 mg Thiamine-HCl  
 1 mg Pyridoxine-HCl  
 1 mg Nicotinic acid  
 100 mg Myo-inositol  
 25 30 g Glucose  
 0.9 g MgCl<sub>2</sub>·6H<sub>2</sub>O  
 2.0 g gellan gum (Phytigel™, Sigma)  
 pH 6.5

## (7) Young plant growing medium

30 S&H medium Macro and Micro elements (Stewart and Hsu, 1977)

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10 mg Thiamine-HCl  
 1 mg Pyridoxine-HCl  
 1 mg Nicotinic acid  
 100 mg Myo-inositol  
 5 10 g Glucose  
 10 g Sucrose  
 0.9 g  $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$   
 2.0 g gellan gum (Phytigel™, Sigma)  
 pH 7.0

10 The following Examples are intended to illustrate the present invention, and not in any way to limit its scope, which is solely defined by the claims.

Example 1: Agrobacterium strain and plasmids

15 *A. tumefaciens* strain LBA 4404 (pBI121GFP) was used for transformation of cotton petiole and young stem. The physical map of pBI121GFP is shown in Fig.1, which contains GFP as a reporter gene and NPTII gene (encoding neomycin phosphotransferase) as a selectable marker. The GFP and NPTII genes are under the control  
 20 of CaMV 35S promoter and nos promoter respectively. For construction of pBI121GFP, a 720 bp XbaI-SstI fragment of GFP gene from the pGFP2 plasmid (from Dr. N. H. Chua, Rockefeller University, New York) was cloned into the same sites in plasmid vector pBI121  
 25 (Clontech) to replace the GUS gene. The pBI121GFP plasmid was introduced into *A. tumefaciens* LBA 4404 by electroporation.

Example 2: Plant material



Upland cotton varieties Coker 312 from the U.S.A. and Si-Mian 3 from Shanxi Cotton Research Institute in China were used in the experiments.

5 Tender petioles were collected from plants 8-12 weeks old grown in a greenhouse with low light conditions. The petioles were surface-sterilized with 70% ethanol for a few seconds, followed by 20% bleach solution (Clorox Co. USA, 1% available chlorine) for 20 min. After rinsing five times in sterilized water, the  
10 petioles were pre-cultured in MS medium for 3 days.

### Example 3: Plant transformation

A single colony of *A. tumefaciens* strain LBA 4404 (pBI121GFP) was inoculated in liquid LB medium with 50 mg/L Rifampicin, 50 mg/L kanamycin and 100 mg/L  
15 streptomycin. The bacteria was grown overnight at 28°C in a shaker of 200 rpm. The bacterium cultures were diluted using liquid MS medium to OD600 = 0.3.

The petiole and young stem were cut into about 2 cm long segments. The segments were soaked in the  
20 diluted bacterium suspension for 5 min, then transferred onto plastic plates (100 x 25 mm) containing a filter paper soaked in 50 ml of co-culture medium. The plates were kept in an incubator of 24°C under continuous light for 48 hrs. The co-cultured  
25 explants were transferred onto MMS1 medium and incubated at 28°C with 16 hrs light ( $60-90 \mu\text{Em}^{-2}\text{s}^{-1}$ ) and 8 hrs dark per day. After 2-4 weeks calli were initiated at the cut ends of petiole segments. After 4-6 weeks kanamycin resistant calli had appeared, and  
30 the number of calli were counted and the expression of GFP gene was examined.

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Under the fluorescence microscope, the untransformed control callus appeared red in colour, while the transformed callus expressing GFP gene displayed distinct green fluorescence. A total of 113 putative transformed calli were examined for GFP activity, the transformation frequency of GFP gene was 39.8% (Table 1). When petioles from cotton variety Si-Mian 3 were used for transformation, 11 calli were found GFP positive from 26 calli tested, transformation efficiency was 42.3%.

Table 1: Transformation frequencies of petioles from cotton Coker 312 and Si-Mian 3

Varieties	Number of calli tested	Number of GFP positive calli	GFP gene transformation frequency (%)
Coker 312	113	45	39.8
Si-Mian 3	26	11	42.3

Example 4: Induction of somatic embryogenesis and plant regeneration

The calli with vigorous growth and strong expression of GFP were selected and transferred into liquid MMS2 medium for suspension culture for 2 weeks. Friable cream-colored granular calli were selected and transferred to semi-solid differential medium, MMS2. After about 2 months a large number of embryoids were produced. Cytoplasmic dense embryogenic structures were gradually developed and large embryos were produced on the medium within 1-2 month. A short time of suspension culture treatment was very important, not only for high frequencies of embryogenesis induction, but also for production of embryoids of good quality.

Expression of GFP gene was checked again and all were GFP positive.

The embryoids and embryogenic calli with strong GFP activity were transferred onto the MMS3 medium.

5 After 1-2 months the plantlets that were about 1-2 cm in height with 1-2 true leaves and good root development were transferred to the Young Plant Growing Medium for about one month. About one month later, young plants with 6-8 leaves and about 10-15 cm in  
10 height were potted in soil and move to the glasshouse. All 30 potted transgenic plants survived and were found expressing GFP protein. The total time required to obtain transgenic plantlets using was under 7 months, and plantlets were reading for potting out in the  
15 greenhouse in about 2 additional months (see Table 2).

Table 2: The time frame from transformation of petiole segments to plant regeneration (Coker 312)

Transform- ation	Callus obtained	Embryos appeared	Regener- ation	Plants planted to green house	Flowering
10/4/98	26/5/98	29/7/98	1/11/98	31/12/98	14/2/99

#### Example 5: Detection of GFP Protein Activity

The expression of GFP protein activity was detected using a Leica MZ FLIII Fluorescence stereo microscope with a 480/40 nM excitation filter and a 510  
25 nM barrier filter.

Green fluorescence of GFP gene can be easily distinguished in the transformed callus, embryoids, and young plantlets, with the untransformed control appeared red in colour under the fluorescence Stereo  
30 microscope. The exceptions were the untransformed

roots, which appeared dim green under the fluorescence microscope, probably due to some chromophorous chemicals accumulated in roots. But the roots with GFP activity could still be identified because the green fluorescence produced by GFP protein was brighter and appeared more uniform. Under the blue light produced by the fluorescence stereo microscope, red fluorescence is clearly visible in untransformed green plant tissues that are enriched with chlorophyll such as leaf and stem. In GFP-positive green plant tissues, yellow fluorescence also was detected because of the overlapping of red and green fluorescence. However, the expression of GFP gene in petal and anther was poorer in comparison to that in other parts of plant.

#### 15 Example 6: Analysis of Transgenic Plants

Genomic DNA from putatively transformed lines and non-transformed control plants was purified according to Paterson et al. (1993). After digestion with *EcoRI*, which cuts in-between left border of T-DNA and Nos-3' terminator of the chimerical GFP gene (Fig. 1), DNA was separated on a 0.8% TAE agarose gel and transferred to Hybond-N membrane according to manufacturer's instructions. DNA was fixed to the membrane by UV crossing linking before hybridizing to the DIG labeled coding region of the GFP gene. Hybridized probe was detected with anti-DIG-AP conjugate according to manufacturer's instructions (BOEHRINGER MANNHEIM).

The genomic DNA samples from 11 randomly selected transgenic lines and 1 untransformed control plant were analyzed Southern hybridization, using the coding region of GFP gene as the hybridization probe. The data indicate that 7 out of 11 lines have a single

copy, 3 lines have 2 copies, and 1 line has 6 copies of  
T-DNA insertion. The high percentage of transgenic  
lines with a single copy of T-DNA insertion suggests  
that this transformation protocol has less risk of gene  
5 silencing and undesirable insertion mutants.

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We claim:

1. A method for producing a transgenic cotton plant comprising the steps of:
  - (a) obtaining cotton petiole explants,
  - (b) exposing the petiole explants to a culture of *Agrobacterium tumefaciens* that harbors a vector comprising an exogenous gene and a selectable marker, the *Agrobacterium* being capable of effecting the stable transfer of the exogenous gene and selection agent resistance gene to the genome of the cells of the petiole explant,
  - (c) culturing the petiole explants to induce callus formation,
  - (d) selecting transformed callus that expresses the exogenous gene,
  - (e) culturing the selected callus in suspension culture to induce formation of embryoids,
  - (f) regenerating the embryoids into whole transgenic cotton plants.
2. The method of claim 1 wherein the petiole explants are pre-cultured for a period of time prior to exposure to the culture of *Agrobacterium tumefaciens*.
3. The method of claims 1 wherein the culture media used in steps (b)-(e) have glucose as the sole carbon source.
4. The method of claim 3 wherein the glucose is in an amount of about 10 g/l to about 50 g/l.

5. The method of claim 4 wherein the glucose is in an amount of about 30 g/l.
6. The method of claim 1 wherein the culture media used in steps (b) and (d)-(f) do not contain hormones.
7. The method of claim 1 wherein embryoid germination is carried out in a medium having a source of nitrogen selected from the group consisting of asparagine, glutamine or both asparagine and glutamine.
8. The method of claim 7 wherein the source of nitrogen is in an amount of about 700 mg/l to about 5 g/l.
9. The method of claim 8 wherein the source of nitrogen is in an amount of about 3.8 g/l.
10. The method of claim 7 wherein the source of nitrogen is both asparagine and glutamine, and the asparagine is in an amount of about 200 mg/l to about 1 g/l and the glutamine is in an amount of about 500 mg/l to about 2 g/l.
11. The method of claim 10 wherein the asparagine is in an amount of about 500 mg/l and the glutamine is in an amount of about 1 g/l.
12. The method of claim 1 wherein the suspension culture of step (e) has a duration of less than about 20 days.

13. The method of claim 12 wherein the suspension culture of step (e) has a duration of about 10 days to about 20 days.
- 5 14. The method of claim 13 wherein the suspension culture of step (e) has a duration of about 14 days.
15. The method of claim 1 wherein step (c) is carried out in the presence of low concentration of one or more hormones.
- 10 16. The method of claim 15 wherein the concentration of any one hormone ranges from 0 to about 1 mg/l.
- 15 17. The method of claim 15 wherein step (c) is carried out in the presence of 2,4-dichlorophenoxyacetic acid in a concentration ranging from 0 to about 0.5 mg/l and kinetin in concentration ranging from 0 to about 1 mg/l.
- 20 18. The method of claim 17 wherein the 2,4-dichlorophenoxyacetic acid is in a concentration of about 0.05 mg/l and the kinetin is in a concentration of about 0.1 mg/l.

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## Abstract

A method is disclosed for producing a transgenic cotton plant by *Agrobacterium*-mediated transformation of petiole tissue. The method comprises the steps of

5 (a) obtaining cotton petiole explants, (b) exposing the petiole explants to a culture of *Agrobacterium tumefaciens* that harbors a vector comprising an exogenous gene and a selectable marker, the

10 *Agrobacterium* being capable of effecting the stable transfer of the exogenous gene and selection agent resistance gene to the genome of the cells of the petiole explant, (c) culturing the petiole explants to induce callus formation, (d) selecting transformed

15 callus that expresses the exogenous gene, (e) culturing the selected callus in suspension culture to induce formation of embryoids, (f) regenerating the embryoids into whole transgenic cotton plants.

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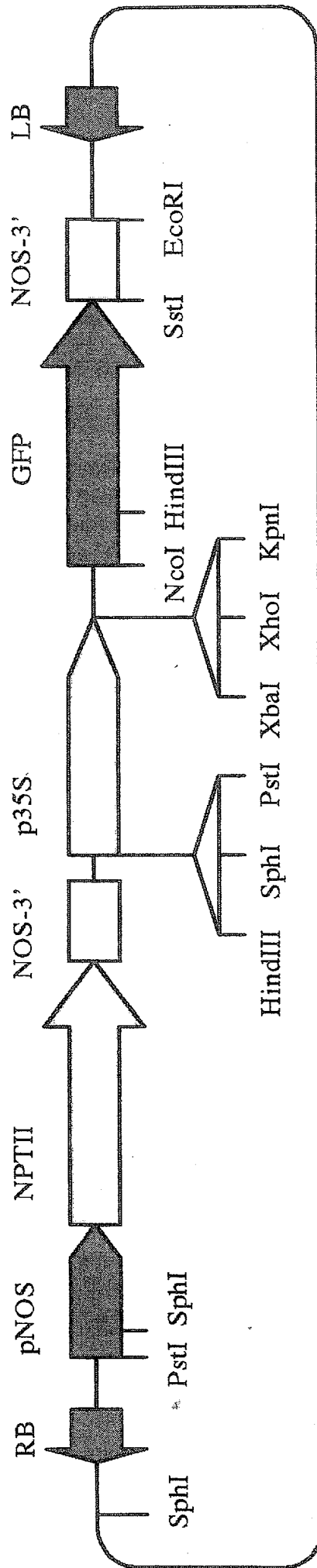


Fig. 1.

<b>DECLARATION AND POWER OF ATTORNEY FOR UTILITY OR DESIGN PATENT APPLICATION</b> (37 CFR 1.63)	Attorney Docket No.	2577-124
	First Named Inventor	Chen Zhi XIAN et al.
	COMPLETE IF KNOWN	
	Application Number	
	Filing Date	
	Group Art Unit	
	Examiner Name	

☐ Declaration  
Submitted  
with Initial  
Filing

☐ Declaration  
Submitted  
after Initial  
Filing

As a below named inventor, I hereby declare that:

My residence, mailing address, and citizenship are as stated below next to name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled: HIGH-EFFICIENCY AGROBACTERIUM-MEDIATED TRANSFORMATION OF COTTON USING PETIOLE EXPLANTS the specification of which was filed on June 11, 1998 as PCT International Application Number PCT/SG99/00058 and was amended on ..

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment specifically referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR 1.56, including for continuation-in-part applications, material information which became available between the filing date of the prior application and the national or PCT international filing date of the continuation-in-part application.

I hereby claim foreign priority benefits under 35 U.S.C. 119(a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or any PCT international application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application Numbers	Country	Foreign Filing Date (MM/DD/YYYY)	Priority Not Claimed	Certified Copy Attached?	
			<input type="checkbox"/>	YES <input type="checkbox"/>	NO <input type="checkbox"/>

I hereby claim the benefit under 35 U.S.C. 119(e) of any United States provisional application(s) listed below.

Application Number(s)	Filing Date (MM/DD/YYYY)

I or we hereby appoint the registered practitioner(s) associated with Customer No. 6449 to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith. Direct all correspondence to Customer Number 6449.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under 18 U.S.C. 1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.



1-00

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